

WO0008157

Publication Title:

HUMAN ANION TRANSPORTER GENES ATNOV

Abstract:

126b Abstract of WO0008157

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ATTORNEY DOCKET NUMBER: 9301-235-999
SERIAL NUMBER: 10/813,506
REFERENCE: **B40**

PCTWORLD INTELLECTUAL PROPERTY ORGANIZATION
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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12N 15/12, 5/10, C12Q 1/68, A01K 67/00, C12N 15/00, C07K 16/28, 14/705		A2	(11) International Publication Number: WO 00/08157 (43) International Publication Date: 17 February 2000 (17.02.00)
(21) International Application Number: PCT/US99/17823 (22) International Filing Date: 6 August 1999 (06.08.99) (30) Priority Data: 60/095,835 7 August 1998 (07.08.98) US (71) Applicant (for all designated States except US): AXYS PHARMACEUTICALS, INC. [US/US]; 180 Kimball Way, South San Francisco, CA 94080 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): LAUBERT, Boris [US/US]; 4550 Bancroft Street #4, San Diego, CA 92116 (US). CARDOSO, Gizela [US/US]; 721 North Main Street, Brockton, Maine 02401 (US). HU, Ping [US/US]; 5807 Folkstone Road, Bethesda, MD 20817 (US). MILLER, Andrew, P. [US/US]; 3271 Countryside Drive, San Mateo, CA 94403 (US). BUCKLER, Alan, J. [US/US]; 2315 Lagoon View Drive, Cardiff, CA 92007 (US). (74) Agent: SHERWOOD, Pamela, J.; Bozicevic, Field & Francis LLP, Suite 200, 285 Hamilton Avenue, Palo Alto, CA 94301 (US).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>	
(54) Title: HUMAN ANION TRANSPORTER GENES ATNOV			
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HUMAN ANION TRANSPORTER GENES

INTRODUCTION

Background

5 Endo- and xenobiotics are typically cleared from mammals via the liver, the primary site of drug metabolizing enzymes. Charged compounds, either endogenously or exogenously derived, are taken up by hepatocytes across the basolateral membrane, appropriately metabolized by the liver enzymes, trafficked through the cell, and then excreted across the canalicular membrane into the bile. These four steps are important in
10 determining a patient's response to pharmaceutical agents.

 Generation of bile flow is a regulated, ATP-dependent process and depends on the coordinated action of a number of transporter proteins in the sinusoidal and canalicular domains of the hepatocyte. Dysfunction of any of these proteins leads to retention of substrates, with conjugated hyperbilirubinemia or cholestasis as a result. In recent years
15 many of the transport proteins involved in bile formation have been identified, cloned, and functionally characterized. The hepatocyte sinusoidal membrane contains transport proteins for the hepatic uptake of organic anions and cations and for the uptake of bile acids.

 The Na⁺-independent organic anion transporter, OATP, resides on the basolateral surface of hepatocytes and mediates the uptake of a large number of amphipathic
20 substrates, such as bromosulfophalein, bile acids, estrogen conjugates, neutral steroids, organic cations, cardiac glycosides, and peptidomimetic drugs. The human organic anion transporter, OATP, is expressed in multiple tissues, including brain, lung, liver, kidney, and testes, while a rat homolog of OATP is expressed only in liver and kidney (Bergwerk et al. (1996) Am. J. Physiol. 271: G231-G238). A prostaglandin transporter, hPGT, which shares
25 significant homology with these organic anion transporters, is abundantly expressed (Lu et al. (1996) J. Clin. Invest. 98: 1142-1149).

 Variations in transporter sequences may alter the kinetic properties of the protein. For example, inefficient clearance of substrates would result in an increased biological half-life, where drugs have an increased half-life and drug levels approach or reach toxic
30 thresholds. Alternatively, over-efficient clearance of substrates could reduce the biological effectiveness of a drug. The identification of novel genes within these pathways provides additional targets for pharmacogenetic analysis, as well as a more thorough understanding of the biological process of drug clearance.

Relevant Literature

The molecular and functional characterization of an organic anion transporting polypeptide cloned from human liver, OATP, is described by Kullak-Ublick et al. (1995) Gastroenterology 109:1274-1282. Other cloned transporter genes are described by Noe et al. (1997) Proc. Natl. Acad. Sci. 94:10346-10350; and Jacquemin et al. (1994) Proc. Natl. Acad. Sci. 91:133-137.

The role of organic cation transporters in intestine, kidney, liver, and brain is reviewed by Koepsell (1998) Annu Rev Physiol 60:243-266. Canalicular multispecific organic anion transporter and the disposal of endo- and xenobiotics is reviewed by Elferink and Jansen (1994) Pharmac. Ther. 64:77-97.

Public EST sequences having sequence similarity with ATnov nucleic acids include: Genbank accessions nos. N49902 (ATnov2); N50005 (ATnov2); H62927 (ATnov3); H62893 (ATnov3); R29414 (ATnov3); AA382692 (ATnov3); T73863 (ATnov3); T74263 (ATnov3); T55488 (ATnov3).

SUMMARY OF THE INVENTION

Isolated nucleotide compositions and sequences are provided for ATnov genes. The ATnov nucleic acid compositions find use in identifying homologous or related genes; in producing compositions that modulate the expression or function of its encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as treatment of anion transporter defects, identification of cell type based on expression, and the like.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Nucleic acid compositions encoding ATnov anion transporters are provided. They are used in identifying homologous or related genes; in producing compositions that modulate the expression or function of the encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. The ATnov gene products are members of the anion transporter gene family, and have high degrees of homology at the amino acid level with known anion transporters.

CHARACTERIZATION OF ATNOV

The sequence data predict that the provided ATnov genes encode anion transporters. Characterization of organic ion transport across the cell membrane, in terms of substrates, binding and transport kinetics, is an important aspect of ATnov biology. A
5 substrate, as used herein, is a chemical entity that is transported by an ATnov polypeptide, usually under normal physiological conditions. Substrates can be either endogenous substrates, i.e. substrates normally found within the natural environment, such as bile salts, or exogenous, i.e. substrates that are not normally found within the natural environment.

Substrate screening assays are used to determine the kinetics of a ATnov protein or
10 peptide fragment on a substrate. Many suitable assays are known in the art, including the use of primary or cultured cells, genetically modified cells (e.g., where DNA encoding the ATnov polymorphism to be studied is introduced into the cell within an artificial construct), cell-free systems, e.g. recombinantly produced enzymes in a suitable buffer, or in animals, including human clinical trials (see, e.g. (1995) Burchell et al. Life Sci. 57:1819-1831,
15 specifically incorporated herein by reference). Where genetically modified cells are used, since most cell lines do not express ATnov activity (liver cells lines being the exception), introduction of artificial construct for expression of the ATnov polymorphism into many human and non-human cell lines does not require additional modification of the host to inactivate endogenous ATnov expression/activity. Clinical trials may monitor serum, urine,
20 etc. levels of the substrate or its metabolite(s).

Full length ion transporter cDNAs may be combined with proper vectors to form expression constructs of each individual transporter. Functional analyses of expressed transporters can be performed in heterologous systems, or by expression in mammalian cell lines. For expression analyses in heterologous systems such as *Xenopus* oocytes, synthetic
25 mRNA is made through in vitro transcription of each transporter construct. mRNA is then injected into prepared oocytes and the cells allowed to express the transporter for several days. Candidate substrates may be labeled to provide a means of following movement across the membrane. Similarly, the requirements of a transporter for ATP, Na⁺, etc. may be assessed. For an example of these techniques, see Kullak-Ublick et al. (1997)
30 Gastroenterology 113(4):1295-1305.

Heterologous or mammalian cell lines expressing the novel transporters can be used to characterize small molecules and drugs that interact with the transporter. The same

experiments can be used to assay for novel compounds that interact with the expressed transporters.

ATNOV NUCLEIC ACID COMPOSITIONS

5 As used herein, the term "ATnov" is generically used to refer to any one of the provided nucleotide sequences as set forth in the SEQLIST. Of particular interest are the sequences, including polymorphisms, of ATnov3.1 and ATnov3.2. These sequences are provided as SEQ ID NO:3 (ATnov3.1), SEQ ID NO:5 (ATnov3.1), SEQ ID NO:7 (ATnov3.2) and SEQ ID NO:9 (ATnov3.2). The encoded polypeptides are provided as SEQ ID NO:4,
10 6, 8 and 10, respectively. The polymorphic variants are set forth in the sequences listings. These include a G or A polymorphism at nucleotide 487, resulting in an amino acid change of asp to asn. There is a polymorphism of C or T at nucleotide 670, which is silent with respect to the encoded polypeptide. A frameshift variant is found in the poly T stretch between positions 1705 and 1710, where the sequence contains either 5T or 6T. The 5T
15 polymorphism results in a truncated polypeptide product of 542 amino acids (SEQ ID NO:4 and SEQ ID NO:8), while the 6T polymorphism encodes the full-length protein of 591 amino acids.

Also of interest are the genetic sequences of SEQ ID NO:1 (ATnov1) and SEQ ID NO:2 (ATnov2).

20 Where a specific ATnov sequence is intended, the numerical designation will be added. Nucleic acids encoding ATnov anion transporters may be cDNA or genomic DNA or a fragment thereof. The term "ATnov gene" shall be intended to mean the open reading frame encoding any of the provided ATnov polypeptides, introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20
25 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

Novel nucleic acid compositions of the invention of particular interest comprise a sequence set forth in SEQ ID NO:1, 2, 3, 5, 7, 9 or an identifying sequence thereof. An
30 "identifying sequence" is a contiguous sequence of residues at least about 10 nt to about 20 nt in length, usually at least about 50 nt to about 100 nt in length, that uniquely identifies a nucleic acid sequence, e.g., exhibits less than 90%, usually less than about 80% to about 85% sequence identity to any contiguous nucleotide sequence of more than about 20 nt.

Thus, the subject novel nucleic acid compositions include full length cDNAs or mRNAs that encompass an identifying sequence of contiguous nucleotides from SEQ ID NO:1, 2, 3, 5, 7, 9.

The nucleic acids of the invention also include nucleic acids having sequence
5 similarity or sequence identity. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 10XSSC (0.9 M NaCl/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC. Sequence identity can be determined by hybridization under stringent conditions, for example, at 50°C or higher and 0.1XSSC (9 mM NaCl/0.9 mM sodium citrate).
10 Hybridization methods and conditions are well known in the art, see U.S. Patent No. 5,707,829. Nucleic acids that are substantially identical to the provided nucleic acid sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided nucleic acid sequences (SEQ ID NO:1, 2, 3, 5, 7, 9) under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate
15 homologous or related genes. The source of homologous genes can be any species.

Preferably, hybridization is performed using at least 15 contiguous nucleotides of SEQ ID NO:1, 2, 3, 5, 7, 9. The probe will preferentially hybridize with a nucleic acid or mRNA comprising the complementary sequence, allowing the identification and retrieval of the nucleic acids of the biological material that uniquely hybridize to the selected probe.
20 Probes of more than 15 nucleotides can be used, e.g. probes of from about 18 nucleotides to not more than about 100 nucleotides, but 15 nucleotides generally represents sufficient sequence for unique identification.

The nucleic acids of the invention also include naturally occurring variants of the nucleotide sequences, e.g. degenerate variants, allelic variants, etc. Variants of the nucleic
25 acids of the invention are identified by hybridization of putative variants with nucleotide sequences disclosed herein, preferably by hybridization under stringent conditions. For example, by using appropriate wash conditions, variants of the nucleic acids of the invention can be identified where the allelic variant exhibits at most about 25-30% base pair mismatches relative to the selected nucleic acid probe. In general, allelic variants contain
30 5-25% base pair mismatches, and can contain as little as even 2-5%, or 1-2% base pair mismatches, as well as a single base-pair mismatch.

The invention also encompasses homologs corresponding to the nucleic acids of SEQ ID NO:1, 2, 3, 5, 7, 9, where the source of homologous genes can be any related

species within the same genus or group. Within a group, homologs have substantial sequence similarity, e.g. at least 75% sequence identity, usually at least 90%, more usually at least 95% between nucleotide sequences. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. A reference sequence will usually be at least about 18 contiguous nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al., J. Mol. Biol. (1990) 215:403-10.

In general, variants of the invention have a sequence identity greater than at least about 65%, preferably at least about 75%, more preferably at least about 85%, and can be greater than at least about 90% or more as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular). For the purposes of this invention, a preferred method of calculating percent identity is the Smith-Waterman algorithm, using the following. Global DNA sequence identity must be greater than 65% as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular) using an affine gap search with the following search parameters: gap open penalty, 12; and gap extension penalty, 1.

ATnov polymorphic sequences. It has been found that specific sites in the ATnov gene sequence are polymorphic, i.e. within a population, more than one nucleotide (G, A, T, C) is found at a specific position. Polymorphisms may provide functional differences in the genetic sequence, through changes in the encoded polypeptide, changes in mRNA stability, binding of transcriptional and translation factors to the DNA or RNA, and the like.

The polymorphisms are also used as single nucleotide polymorphisms to detect association with, or genetic linkage to phenotypic variation in activity and expression of ATnov.

SNPs are generally biallelic systems, that is, there are two alleles that an individual may have for any particular marker. SNPs, found approximately every kilobase, offer the potential for generating very high density genetic maps, which will be extremely useful for developing haplotyping systems for genes or regions of interest, and because of the nature of SNPs, they may in fact be the polymorphisms associated with the disease phenotypes under study. The low mutation rate of SNPs also makes them excellent markers for studying complex genetic traits.

Single nucleotide polymorphisms are provided in the ATnov3 sequence listing. The provided sequences also encompass the complementary sequence corresponding to any of the provided polymorphisms.

5 In order to provide an unambiguous identification of the specific site of a polymorphism, sequences flanking the polymorphic site are included in a probe for the region. It will be understood that there is no special significance to the length of non-polymorphic flanking sequence that is included, except to aid in positioning the polymorphism in the genomic sequence.

10 For screening purposes, hybridization probes of the polymorphic sequences may be used where both forms are present, either in separate reactions, spatially separated on a solid phase matrix, or labeled such that they can be distinguished from each other. Assays may utilize nucleic acids that hybridize to one or more of the described polymorphisms.

15 An array may include all or a subset of the ATnov3 polymorphisms. One or both polymorphic forms may be present in the array. Usually such an array will include at least 2 different polymorphic sequences, i.e. polymorphisms located at unique positions within the locus, and may include as many all of the provided polymorphisms. Arrays of interest may further comprise sequences, including polymorphisms, of other genetic sequences, particularly other sequences of interest for pharmacogenetic screening. The oligonucleotide sequence on the array will usually be at least about 12 nt in length, may be the length of the
20 provided polymorphic sequences, or may extend into the flanking regions to generate fragments of 100 to 200 nt in length. For examples of arrays, see Ramsay (1998) Nat. Biotech. 16:40-44; Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

25 The subject nucleic acids can be cDNAs or genomic DNAs, as well as fragments thereof, particularly fragments that encode a biologically active gene product and/or are useful in the methods disclosed herein. The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when
30 present, being removed by nuclear RNA splicing, to create a continuous open reading frame encoding a polypeptide of the invention.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the

introns that are normally present in a native chromosome. It can further include the 3' and 5' untranslated regions found in the mature mRNA. It can further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' and 3' end of the transcribed region. The genomic DNA can be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' and 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for expression.

The nucleic acid compositions of the subject invention can encode all or a part of the subject polypeptides. Double or single stranded fragments can be obtained from the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. Isolated nucleic acids and nucleic acid fragments of the invention comprise at least about 15 up to about 100 contiguous nucleotides, or up to the complete sequence provided in SEQ ID NO:1, 2, 3, 5, 7 or 9. For the most part, fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and up to at least about 50 contiguous nt in length or more.

Probes specific to the nucleic acids of the invention can be generated using the nucleic acid sequences disclosed in SEQ ID NO:1, 2, 3, 5, 7 or 9 and the fragments as described above. The probes can be synthesized chemically or can be generated from longer nucleic acids using restriction enzymes. The probes can be labeled, for example, with a radioactive, biotinylated, or fluorescent tag. Preferably, probes are designed based upon an identifying sequence of a nucleic acid of one of SEQ ID NO:1, 2, 3, 5, 7 or 9. More preferably, probes are designed based on a contiguous sequence of one of the subject nucleic acids that remain unmasked following application of a masking program for masking low complexity (e.g., XBLAST) to the sequence., i.e. one would select an unmasked region, as indicated by the nucleic acids outside the poly-n stretches of the masked sequence produced by the masking program.

The nucleic acids of the subject invention are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the nucleic acids, either as DNA or RNA, will be obtained substantially free of other naturally-occurring nucleic acid sequences, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", e.g., flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

The nucleic acids of the invention can be provided as a linear molecule or within a circular molecule. They can be provided within autonomously replicating molecules (vectors) or within molecules without replication sequences. They can be regulated by their own or by other regulatory sequences, as is known in the art. The nucleic acids of the invention can be introduced into suitable host cells using a variety of techniques which are available in the art, such as transferrin polycation-mediated DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated DNA transfer, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, gene gun, calcium phosphate-mediated transfection, and the like.

The subject nucleic acid compositions can be used to, for example, produce polypeptides, as probes for the detection of mRNA of the invention in biological samples (e.g., extracts of cells) to generate additional copies of the nucleic acids, to generate ribozymes or antisense oligonucleotides, and as single stranded DNA probes or as triple-strand forming oligonucleotides. The probes described herein can be used to, for example, determine the presence or absence of the nucleic acid sequences as shown in SEQ ID NO:1, 2, 3, 5, 7 or 9 or variants thereof in a sample.

The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where ATnov genes are expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms in the promoter regions are useful for determining natural variations in expression, particularly those that may be associated with disease.

Alternatively, mutations may be introduced into the promoter regions to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) Mol Med 1: 194-205; Mortlock et al. (1996) Genome Res. 6: 327-33; and Joulin and Richard-Foy (1995) Eur J Biochem 232: 620-626.

The regulatory sequences may be used to identify cis acting sequences required for transcriptional or translational regulation of ATnov expression, especially in different tissues or stages of development, and to identify cis acting sequences and trans acting factors that regulate or mediate ATnov expression. Such transcription or translational control regions

may be operably linked to a ATnov gene in order to promote expression of wild type or altered ATnov or other proteins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

Double or single stranded fragments may be obtained of the DNA sequence by
5 chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. For the most part, DNA fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and may be at least about 50 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 100 nt are useful for production of the encoded
10 polypeptide. For use in amplification reactions, such as PCR, a pair of primers will be used. The exact composition of the primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for
15 the selection of primer sequences are generally known, and are available in commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

The DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide
20 sequences, as genomic DNA or RNA, is well established in the literature and does not require elaboration here. DNA or mRNA is isolated from a cell sample. The mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, the mRNA sample is separated by gel electrophoresis,
25 transferred to a suitable support, e.g. nitrocellulose, nylon, etc., and then probed with a fragment of the subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, in situ hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to the subject sequence is indicative of ATnov gene expression in the sample.

30 The sequence of an ATnov gene, including flanking promoter regions and coding regions, may be mutated in various ways known in the art to generate targeted changes in promoter strength, sequence of the encoded protein, etc. The DNA sequence or protein product of such a mutation will usually be substantially similar to the sequences provided

herein, i.e. will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions. Deletions may further include larger changes, such as deletions of a domain or exon. Other modifications of interest include
5 epitope tagging, e.g. with the FLAG system, HA, etc. For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used.

Techniques for in vitro mutagenesis of cloned genes are known. Examples of protocols for site specific mutagenesis may be found in Gustin et al., *Biotechniques* 14:22 (1993); Barany, *Gene* 37:111-23 (1985); Colicelli et al., *Mol Gen Genet* 199:537-9 (1985);
10 and Prentki et al., *Gene* 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, CSH Press 1989, pp. 15.3-15.108; Weiner et al., *Gene* 126:35-41 (1993); Sayers et al., *Biotechniques* 13:592-6 (1992); Jones and Winistorfer, *Biotechniques* 12:528-30 (1992); Barton et al., *Nucleic Acids Res* 18:7349-55 (1990); Marotti and Tomich, *Gene Anal Tech* 6:67-70 (1989); and Zhu, *Anal*
15 *Biochem* 177:120-4 (1989). Such mutated genes may be used to study structure-function relationships of ATnov polypeptides, or to alter properties of the protein that affect its function or regulation.

Genetic polymorphisms, either naturally occurring or introduced as described above, are useful in screening for altered transport or metabolism of ATnov substrates. For
20 example, variant alleles may affect the pharmacokinetic parameters of substrates. A drug's volume of distribution, clearance, and the derived parameter, half-life, are particularly important, as they determine the degree of fluctuation between a maximum and minimum plasma concentration during a dosage interval, the magnitude of steady state concentration and the time to reach steady state plasma concentration upon chronic dosing. Parameters
25 derived from in vivo drug administration are useful in determining the clinical effect of a particular ATnov genotype.

ATNOV POLYPEPTIDES

The subject gene may be employed for producing all or portions of ATnov
30 polypeptides. Fragments of interest include the glycosylation sites, transmembrane domains, ATP binding regions, the substrate binding sites, etc. Such domains will usually include at least about 20 amino acids of the provided sequence, more usually at least about 50 amino acids, and may include 100 amino acids or more, up to the complete domain.

Binding contacts may be comprised of non-contiguous sequences, which are brought into proximity by the tertiary structure of the protein. The sequence of such fragments may be modified through manipulation of the coding sequence, as described above. Truncations may be performed at the carboxy or amino terminus of the fragment, e.g. to determine the
5 minimum sequence required for biological activity.

A subset of the provided nucleic acid polymorphisms in ATnov3 confer a change in the corresponding amino acid sequence, as previously described. Using the amino acid sequence provided in SEQ ID NO:3 as a reference, the amino acid polymorphisms of the invention include asn→asp, pos. 130; and a frameshift at position 537 resulting in a truncated
10 protein of 542 amino acids. Polypeptides comprising at least one of the provided polymorphisms (ATnov3^v polypeptides) are of interest. The term "ATnov3^v polypeptides" as used herein includes complete ATnov protein forms, e.g. such splicing variants as known in the art, and fragments thereof, which fragments may comprise short polypeptides, epitopes, functional domains; binding sites; etc.; and including fusions of the subject polypeptides to
15 other proteins or parts thereof. Polypeptides will usually be at least about 8 amino acids in length, more usually at least about 12 amino acids in length, and may be 20 amino acids or longer, up to substantially the complete protein.

For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or
20 constitutive, where the coding region is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to an ATnov gene, or may be derived from exogenous sources.

The peptide may be expressed in prokaryotes or eukaryotes in accordance with
25 conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli*, *B. subtilis*, *S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, e.g. COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the ATnov gene in eukaryotic cells, where the ATnov
30 protein will benefit from native folding and post-translational modifications. Small peptides can also be synthesized in the laboratory. Peptides that are subsets of the complete ATnov sequence may be used to identify and investigate parts of the protein important for function, or to raise antibodies directed against these regions.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique. The purified protein will generally be at least about 80% pure, preferably at least about 90% pure, and may be up to and including 100% pure. Pure is intended to mean free of other proteins, as well as cellular debris.

The expressed ATnov polypeptides are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Antibodies may be raised to the wild-type or variant forms of ATnov. Antibodies may be raised to isolated peptides corresponding to these domains, or to the native protein.

Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see Monoclonal Antibodies: A Laboratory Manual, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in *E. coli*, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to in vivo immunization as a method of raising antibodies include binding to phage "display" libraries, usually in conjunction with in vitro affinity maturation.

ATNOV GENOTYPING

The subject nucleic acid and/or polypeptide compositions may be used in genotyping and to screen for the presence of polymorphisms in the sequence, or variation in the expression of the subject genes. Genotyping may be performed to determine whether a particular polymorphisms is associated with a disease state or genetic predisposition to a disease state, particularly diseases associated with liver disorders.

Genotyping may also be performed for pharmacogenetic analysis to assess the association between an individual's genotype and that individual's ability to react to a therapeutic agent. Differences in substrate transport to relevant cells can lead to toxicity or therapeutic failure. Relationships between polymorphisms in transporter expression or
5 specificity can be used to optimize therapeutic dose administration.

ATnov genotyping is performed by DNA or RNA sequence and/or hybridization analysis of any convenient sample from a patient, e.g. biopsy material, blood sample, scrapings from cheek, etc. A nucleic acid sample from an individual is analyzed for the presence of polymorphisms in ATnov, particularly those that affect the activity,
10 responsiveness or expression of ATnov. Specific sequences of interest include any polymorphism that leads to changes in basal expression in one or more tissues, to changes in the modulation of ATnov expression, or alterations in ATnov specificity and/or activity.

The effect of a polymorphism in ATnov gene sequence on the response to a particular agent may be determined by in vitro or in vivo assays. Such assays may include
15 monitoring during clinical trials, testing on genetically defined cell lines, etc. The response of an individual to the agent can then be predicted by determining the ATnov genotype with respect to the polymorphism. Where there is a differential distribution of a polymorphism by racial background, guidelines for drug administration can be generally tailored to a particular ethnic group.

20 Biochemical studies may be performed to determine whether a sequence polymorphism in a ATnov coding region or control regions is associated with disease. Disease associated polymorphisms may include deletion or truncation of the gene, mutations that alter expression level, that affect the specificity or transport kinetics of the transporter, etc.

25 A number of methods are available for analyzing nucleic acids for the presence of a specific sequence. Where large amounts of DNA are available, genomic DNA is used directly. Alternatively, the region of interest is cloned into a suitable vector and grown in sufficient quantity for analysis. The nucleic acid may be amplified by conventional techniques, such as the polymerase chain reaction (PCR), to provide sufficient amounts for
30 analysis. The use of the polymerase chain reaction is described in Saiki et al. (1985) Science 239:487, and a review of current techniques may be found in Sambrook et al. Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2-14.33. Amplification may be used to determine whether a polymorphism is present, by using a primer that is

specific for the polymorphism. Alternatively, various methods are known in the art that utilize oligonucleotide ligation as a means of detecting polymorphisms, for examples see Delahunty et al. (1996) Am. J. Hum. Genet. 58:1239-1246.

A detectable label may be included in an amplification reaction. Suitable labels
5 include fluorochromes, e.g. fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'-dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX), 6-carboxy-2',4',7',4,7-hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6-carboxyrhodamine (TAMRA), radioactive labels, e.g. ^{32}P , ^{35}S , ^3H ; etc. The label may be a
10 two stage system, where the amplified DNA is conjugated to biotin, haptens, etc. having a high affinity binding partner, e.g. avidin, specific antibodies, etc., where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

15 The sample nucleic acid, e.g. amplified or cloned fragment, is analyzed by one of a number of methods known in the art. The nucleic acid may be sequenced by dideoxy or other methods. Hybridization with the variant sequence may also be used to determine its presence, by Southern blots, dot blots, etc. The hybridization pattern of a control and variant sequence to an array of oligonucleotide probes immobilised on a solid support, as described
20 in U.S. 5,445,934, or in WO95/35505, may also be used as a means of detecting the presence of variant sequences. Single strand conformational polymorphism (SSCP) analysis, denaturing gradient gel electrophoresis (DGGE), mismatch cleavage detection, and heteroduplex analysis in gel matrices are used to detect conformational changes created by DNA sequence variation as alterations in electrophoretic mobility. Alternatively, where a
25 polymorphism creates or destroys a recognition site for a restriction endonuclease (restriction fragment length polymorphism, RFLP), the sample is digested with that endonuclease, and the products size fractionated to determine whether the fragment was digested. Fractionation is performed by gel or capillary electrophoresis, particularly acrylamide or agarose gels.

30 In one embodiment of the invention, an array of oligonucleotides are provided, where discrete positions on the array are complementary to one or more of the provided polymorphic sequences, e.g. oligonucleotides of at least 12 nt, frequently 20 nt, or larger, and including the sequence flanking the polymorphic position. Such an array may comprise

a series of oligonucleotides, each of which can specifically hybridize to a different polymorphism. For examples of arrays, see Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

5 Screening for polymorphisms in ATnov may be based on the functional or antigenic characteristics of the protein. Protein truncation assays are useful in detecting deletions that may affect the biological activity of the protein. Various immunoassays designed to detect polymorphisms in ATnov proteins may be used in screening. Where many diverse genetic mutations lead to a particular disease phenotype, functional protein assays have proven to
10 be effective screening tools. The activity of the encoded ATnov protein as a anion transporter may be determined by comparison with the wild-type protein.

 Antibodies specific for a ATnov may be used in staining or in immunoassays. Samples, as used herein, include biological fluids such as semen, blood, cerebrospinal fluid, tears, saliva, lymph, dialysis fluid and the like; organ or tissue culture derived fluids; and
15 fluids extracted from physiological tissues. Also included in the term are derivatives and fractions of such fluids. The cells may be dissociated, in the case of solid tissues, or tissue sections may be analyzed. Alternatively a lysate of the cells may be prepared.

 Diagnosis may be performed by a number of methods to determine the absence or presence or altered amounts of normal or abnormal ATnov polypeptides in patient cells. For
20 example, detection may utilize staining of cells or histological sections, performed in accordance with conventional methods. The antibodies of interest are added to the cell sample, and incubated for a period of time sufficient to allow binding to the epitope, usually at least about 10 minutes. The antibody may be labeled with radioisotopes, enzymes, fluorescers, chemiluminescers, or other labels for direct detection. Alternatively, a second
25 stage antibody or reagent is used to amplify the signal. Such reagents are well known in the art. For example, the primary antibody may be conjugated to biotin, with horseradish peroxidase-conjugated avidin added as a second stage reagent. Alternatively, the secondary antibody conjugated to a fluorescent compound, e.g. fluorescein, rhodamine, Texas red, etc. Final detection uses a substrate that undergoes a color change in the
30 presence of the peroxidase. The absence or presence of antibody binding may be determined by various methods, including flow cytometry of dissociated cells, microscopy, radiography, scintillation counting, etc.

MODULATION OF GENE EXPRESSION

The ATnov genes, gene fragments, or the encoded protein or protein fragments are useful in gene therapy to treat disorders associated with ATnov defects. Expression vectors may be used to introduce the ATnov gene into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about one day, more usually for a period of at least about several days to several weeks.

The gene or ATnov protein may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of vesicles. Jet injection may also be used for intramuscular administration, as described by Furth et al. (1992) Anal Biochem 205:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang et al. (1992) Nature 356:152-154), where gold microprojectiles are coated with the ATnov or DNA, then bombarded into skin cells.

Antisense molecules can be used to down-regulate expression of ATnov in cells. The anti-sense reagent may be antisense oligonucleotides (ODN), particularly synthetic ODN having chemical modifications from native nucleic acids, or nucleic acid constructs that express such anti-sense molecules as RNA. The antisense sequence is complementary to the mRNA of the targeted gene, and inhibits expression of the targeted gene products. Antisense molecules inhibit gene expression through various mechanisms, e.g. by reducing the amount of mRNA available for translation, through activation of RNase H, or steric hindrance. One or a combination of antisense molecules may be administered, where a combination may comprise multiple different sequences.

Antisense molecules may be produced by expression of all or a part of the target gene sequence in an appropriate vector, where the transcriptional initiation is oriented such that an antisense strand is produced as an RNA molecule. Alternatively, the antisense molecule is a synthetic oligonucleotide. Antisense oligonucleotides will generally be at least about 7, usually at least about 12, more usually at least about 20 nucleotides in length, and not more than about 500, usually not more than about 50, more usually not more than about

35 nucleotides in length, where the length is governed by efficiency of inhibition, specificity, including absence of cross-reactivity, and the like. It has been found that short oligonucleotides, of from 7 to 8 bases in length, can be strong and selective inhibitors of gene expression (see Wagner et al. (1996) Nature Biotechnology 14:840-844).

5 A specific region or regions of the endogenous sense strand mRNA sequence is chosen to be complemented by the antisense sequence. Selection of a specific sequence for the oligonucleotide may use an empirical method, where several candidate sequences are assayed for inhibition of expression of the target gene in an in vitro or animal model. A combination of sequences may also be used, where several regions of the mRNA sequence
10 are selected for antisense complementation.

 Antisense oligonucleotides may be chemically synthesized by methods known in the art (see Wagner et al. (1993) *supra.* and Milligan et al., *supra.*) Preferred oligonucleotides are chemically modified from the native phosphodiester structure, in order to increase their intracellular stability and binding affinity. A number of such modifications have been
15 described in the literature, which alter the chemistry of the backbone, sugars or heterocyclic bases.

 Among useful changes in the backbone chemistry are phosphorothioates; phosphorodithioates, where both of the non-bridging oxygens are substituted with sulfur; phosphoroamidites; alkyl phosphotriesters and boranophosphates. Achiral phosphate
20 derivatives include 3'-O-5'-S-phosphorothioate, 3'-S-5'-O-phosphorothioate, 3'-CH₂-5'-O-phosphonate and 3'-NH-5'-O-phosphoroamidate. Peptide nucleic acids replace the entire ribose phosphodiester backbone with a peptide linkage. Sugar modifications are also used to enhance stability and affinity. The β -anomer of deoxyribose may be used, where the base is inverted with respect to the natural α -anomer. The 2'-OH of the ribose sugar
25 may be altered to form 2'-O-methyl or 2'-O-allyl sugars, which provides resistance to degradation without comprising affinity. Modification of the heterocyclic bases must maintain proper base pairing. Some useful substitutions include deoxyuridine for deoxythymidine; 5-methyl-2'-deoxycytidine and 5-bromo-2'-deoxycytidine for deoxycytidine. 5-propynyl-2'-deoxyuridine and 5-propynyl-2'-deoxycytidine have been shown to increase affinity and
30 biological activity when substituted for deoxythymidine and deoxycytidine, respectively.

 As an alternative to anti-sense inhibitors, catalytic nucleic acid compounds, e.g. ribozymes, anti-sense conjugates, etc. may be used to inhibit gene expression. Ribozymes may be synthesized in vitro and administered to the patient, or may be encoded on an

expression vector, from which the ribozyme is synthesized in the targeted cell (for example, see International patent application WO 9523225, and Beigelman et al. (1995) Nucl. Acids Res 23:4434-42). Examples of oligonucleotides with catalytic activity are described in WO 9506764. Conjugates of anti-sense ODN with a metal complex, e.g. terpyridyl/Cu(II), capable of mediating mRNA hydrolysis are described in Bashkin et al. (1995) Appl Biochem Biotechnol 54:43-56.

GENETICALLY ALTERED CELL OR ANIMAL MODELS FOR ATNOV FUNCTION

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. Transgenic animals may be made through homologous recombination, where the normal ATnov locus is altered. Alternatively, a nucleic acid construct is randomly integrated into the genome. Vectors for stable integration include plasmids, retroviruses and other animal viruses, YACs, and the like.

The modified cells or animals are useful in the study of ATnov function and regulation. For example, a series of small deletions and/or substitutions may be made in the ATnov gene to determine the role of different transmembrane domains, of ATP catalysis, etc. Of interest are the use of ATnov to construct transgenic animal models where expression of ATnov is specifically reduced or absent. Specific constructs of interest include anti-sense ATnov, which will block ATnov expression, expression of dominant negative ATnov mutations, etc. One may also provide for expression of the ATnov gene or variants thereof in cells or tissues where it is not normally expressed or at abnormal times of development.

DNA constructs for homologous recombination will comprise at least a portion of the ATnov gene with the desired genetic modification, and will include regions of homology to the target locus. DNA constructs for random integration need not include regions of homology to mediate recombination. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are known in the art. For various techniques for transfecting mammalian cells, see Keown et al. (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g. mouse, rat, guinea pig, etc. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES or embryonic cells have been transformed, they may be used to

produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the construct. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting offspring screened for the construct. By providing for a different phenotype of the blastocyst and the genetically modified cells, chimeric progeny can be readily detected.

The chimeric animals are screened for the presence of the modified gene and males and females having the modification are mated to produce homozygous progeny. If the gene alterations cause lethality at some point in development, tissues or organs can be maintained as allogeneic or congenic grafts or transplants, or in in vitro culture. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, etc. The transgenic animals may be used in functional studies, drug screening, etc.

TESTING OF ATNOV FUNCTION and RESPONSES

Anion transporters such as ATnov polypeptides are involved in multiple biologically important processes. Pharmacological agents designed to affect only specific transporter subtypes are of particular interest. The subject polypeptides may be used to test the specificity of novel compounds, and of analogs and derivatives of compounds known to be substrates, or to act on anion transporters.

Drug screening may be performed using an in vitro model, a genetically altered cell or animal, or purified ATnov protein. One can identify ligands or substrates that bind to, modulate or mimic the action of ATnov. Drug screening identifies agents that provide a replacement for ATnov function in abnormal cells. Of particular interest are screening assays for agents that have a low toxicity for human cells. A wide variety of assays may be used for this purpose, including monitoring cellular excitation and conductance, labeled in vitro protein-protein binding assays, electrophoretic mobility shift assays, immunoassays for protein binding, and the like. The purified protein may also be used for determination of

three-dimensional crystal structure, which can be used for modeling intermolecular interactions.

The term "agent" as used herein describes any molecule, e.g. protein or pharmaceutical, with the capability of altering or mimicking the physiological function of
5 ATnov polypeptide. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, though typically they are
10 organic molecules, preferably small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or
15 heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of
20 synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds
25 are readily modified through conventional chemical, physical and biochemical means, and may be used to produce combinatorial libraries. Known pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, amidification, etc. to produce structural analogs.

Where the screening assay is a binding assay, one or more of the molecules may be
30 joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemilumescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific

binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc. that are used to facilitate
5 optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40°C. Incubation periods are selected for
10 optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

The compounds having the desired pharmacological activity may be administered in a physiologically acceptable carrier to a host in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, etc.
15 Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of therapeutically active compound in the formulation may vary from about 0.1-100 wt.%. The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the like. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for
20 oral and topical use can be used to make up compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents.

25

EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (e.g. amounts, temperature, concentrations, etc.) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

Example 1

Three novel members of the OATP gene family, which are expressed in liver tissue, were cloned. These genes were isolated using trapped exons obtained from large-scale exon trapping of chromosome 12. The three anion transporters reported here are 70-80% identical to each other over the predicted protein sequence, and are each 40% identical to the reported OATP protein sequence (Kullack-Ublick et al., (1995) Gastroenterology 109:1274-1282). The chromosomal location of these three anion transporters, along with the mapping of OATP, suggests this gene-family is clustered on 12p12.

Materials and Methods

cDNA Isolation. cDNA clones were isolated using the GeneTrapper system (Gibco-BRL). PCR primers within the trapped exons were used to detect which plasmid cDNA libraries contained the gene of interest. Oligonucleotide probes were designed: (SEQ ID NO:7) C12B_120: GGGGCTCTGATTGATACAACGTG ; (SEQ ID NO:8) C12C_151: ACTGTGGCACACGTGGGTCATGTAGGACAT) and the process proceeded according to the supplied protocol. cDNA clones were sequenced on an ABI 377 according to standard methods. The Primer Island Transposition kit was used according to the supplied protocol. Sequences were analyzed, edited, and assembled using the Sequencher software (Gene Codes).

Radiation Hybrid Mapping. RH mapping was achieved using the Stanford G3 panel DNAs (Research Genetics). DNA was aliquoted into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. 20 µl PCR reactions with standard conditions, 2.5

mM MgCl₂, Taq Gold, and an annealing temperature of 60°C (for ATnov1 and 2) or 55°C (for ATnov3) were used to detect expression. The assays were done in duplicate and results were scored and map positions determined via the RH server at Stanford University <<http://www-shgc.stanford.edu/RH/G3index.html>>.

5

RT-PCR. RT-PCR was utilized to characterize the expression pattern of the novel anion transporters. This approach used RNA from 30 different tissues to generate first strand cDNA. Total RNA was purchased (Clontech, Invitrogen) and used to synthesize first strand cDNA using M-MLV reverse transcriptase and the supplied buffer (Gibco-BRL). The 20 µl reaction contained 5 µg total RNA, 100 ng of random primers, 10 mM DTT, 0.5 mM each dNTP, and an RNase inhibitor (Gibco-BRL). Identical reactions were set up without reverse transcriptase to control for DNA contamination in the RNA samples. The synthesis reaction proceeded for 1 hour at 37°C followed by 10 minutes at 95°C. These cDNAs, along with control cDNA synthesis reactions without reverse transcriptase, were diluted 1:5 and 2 µl of each sample were arrayed into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. The cDNAs were tested with primers with defined expression patterns to verify the presence of amplifiable cDNA from each tissue. Gene-specific primers were used to amplify the cDNAs in 20 µl PCR reactions with standard conditions, 2.5 mM MgCl₂, Taq Gold, and an appropriate annealing temperature.

This approach provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner and provides qualitative analysis of gene expression only. Modifications can be employed, such as the use of internal control primers, limited cycling parameters, and dilution series to convert this to a quantitative experiment.

25 Primers for ATnov1

RH primers (SEQ ID NO:11) CTGCTGCCAACTAACATTGC

(SEQ ID NO:12) CACACACTAACCATGCCTCT

237 bp product

RT-PCR primers (SEQ ID NO:13) TCCAGTCATTGGCTTTGCAC

30 (SEQ ID NO:14) AAGAACCAATAAAGCTGCTTACT

413 bp product

Primers for ATnov2

RH primers (SEQ ID NO:15) GTGTTTGCTAGCCACCTTGA

(SEQ ID NO:16) GGCAACACTTCCTCAAAGTG

196 bp product

RT-PCR primers (SEQ ID NO:17) GATGCTTTCCTCTGTGCAGT
(SEQ ID NO:18) CCTTCAAGCCGAAGAAGGCT

5 259 bp product

Primers for ATnov3

RH primers (SEQ ID NO:19) AGGAGTTCCTGGTCCTTTCA
(SEQ ID NO:20) CAAGCTAGACTTCAGGCCTT

10 137 bp product

RT-PCR primers (SEQ ID NO:21) GAGGAATTCTAGCTCCAATATATT
(SEQ ID NO:22) GTCCTACATGACCCACGTGTG

96 bp product

15 Results

cDNA Isolation. Large-scale exon trapping was completed across a chromosome 12 cosmid library. Approximately 2400 exons were sequenced and analyzed by BLAST algorithms to identify exons with potentially interesting homologies. Two different exons, C12B_120 and C12C_151 were identified that were 87% identical to each other and ~68% identical to a cloned organic anion transporter, OATP (Kullak-Ubrich et al. (1995) Gastroenterology 109: 1274-1282.), at the DNA level and ~78% similar at the amino acid level. Full-length cDNA clones were isolated using GeneTrapper (Gibco-BRL) from a liver cDNA library (Gibco-BRL). The resulting clones, the largest being up to 3.0 kb, were end-sequenced using vector primers. If the end sequences provided insufficient coverage of the cDNA clones, a transposon approach was used to complete the sequence of the cDNA clone.

The cDNA clones isolated with C12B_120 yielded two different sequence contigs, ATnov1 and ATnov2, which were ~89% identical to each other. ATnov2 is identical to C12B_120. cDNA clones isolated with C12C_151 generated a sequence contig, ATnov3, that was ~86% identical to the first two contigs. Conceptual translations yielded predicted proteins of 688-704 amino acids in length. A multiple alignment of these three proteins is shown in figure 1. These genes also show significant homology to a human organic anion

transporter OATP (~40% identity, 60% similarity) and to a human prostaglandin transporter (~32% identity, 51% similarity) over the length of the predicted proteins.

Chromosomal Localization. The exon trapped products used in the cDNA screens
5 were trapped from a chromosome 12 cosmid library, suggesting that at least ATnov2 and
ATnov3 map to chromosome 12. OATP had been previously reported to map to
chromosome 12 (Kullak-Ubrich et al., supra.) Radiation hybrid mapping was used to confirm
the localization of these to chromosome 12, as well as to map them and ATnov1 to a specific
region on the chromosome. The Stanford G3 panel showed linkage of all four of these
10 genes to the marker GATA91H01, which is extrapolated to a cytogenetic location of 12p12.

Expression Analysis. OATP is expressed in multiple tissues, including brain, lung,
liver, kidney, and testes (Kullak-Ubrich et al., supra.) RT-PCR was utilized to characterize
the expression pattern of the novel anion transporters. This approach used RNA from 30
15 different tissues to generate first strand cDNA. These cDNAs were arrayed, along with
control cDNA synthesis reactions without reverse transcriptase, into 96-well trays, dried and
stored until needed. This resource provides for relatively high-throughput analysis of gene
expression in a large set of tissues in a cost-efficient manner. RT-PCR in this fashion allows
for qualitative analysis of gene expression only.

20 PCR was performed on these plates with gene-specific primers for each of the ATnov
genes. ATnov1 is expressed in fetal and adult liver; ATnov2 is expressed in adult liver and
mammary gland; ATnov3 is expressed in fetal liver, adult liver, brain, adipose tissue, skin,
and testes.

25 The predicted positions of transmembrane domains in the ATnov3 polypeptide are
as follows:

	ATnov3
Transmembrane domain 1	29-45
Transmembrane domain 2	94-104
Transmembrane domain 3	169-189
Transmembrane domain 4	207-227
Transmembrane domain 5	259-279
Transmembrane domain 6	336-356

Transmembrane domain 7	376-396
Transmembrane domain 8	410-430
Transmembrane domain 9	481-501
Transmembrane domain 10	537-557
Transmembrane domain 11	581-601
Transmembrane domain 12	627-647

These novel members of the organic anion transporter family are expressed in the liver. Based on homology to another organic anion transporter, they are likely to be present on the basolateral surface of the hepatocytes and mediate the uptake of both xenobiotics and endogenous compounds for metabolism by the cytochrome p450s, glucuronosyl transferases, and other metabolic enzymes known to be present in the liver. The ATnov genes are all expressed in the liver, with ATnov 2 and 3 also being expressed in a limited number of other tissues. The RT-PCR approach described herein has a high level of sensitivity, with the ability to detect a control transcript diluted down to an expression level equivalent to a frequency of 1/10⁷.

The map positions of these anion transporters suggest that they lie adjacent to each other on the proximal short arm of chromosome 12. The anion transporters described herein are only ~89% identical to each other at the DNA level, suggesting that these genes arose via a recombination mechanism, but have since diverged sufficiently such that it is unlikely that these genes are polymorphic within a given population.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid encoding a mammalian ATnov protein.
2. An isolated nucleic acid according to Claim 1, wherein said ATnov protein has
5 the amino acid sequence of SEQ ID NO:4, 6, 8, or 10.
3. An isolated nucleic acid according to Claim 1, wherein said ATnov protein has
an amino acid sequence that is substantially identical to the amino acid sequence of SEQ
ID NO:4, 6, 8, or 10.
10
4. An isolated nucleic acid according to Claim 1, comprising the nucleotide
sequence as set forth in SEQ ID NO:1,2, 3, 5, 7, or 9.
5. An isolated nucleic acid that hybridizes under stringent conditions to the
15 nucleic acid sequence of claim 4.
6. An expression cassette comprising a transcriptional initiation region functional
in an expression host, a nucleic acid having a sequence of the isolated nucleic acid
according to Claim 1 under the transcriptional regulation of said transcriptional initiation
20 region, and a transcriptional termination region functional in said expression host.
7. A cell comprising an expression cassette according to Claim 6 as part of an
extrachromosomal element or integrated into the genome of a host cell as a result of
introduction of said expression cassette into said host cell, and the cellular progeny of said
25 host cell.
8. A method for producing mammalian ATnov protein, said method comprising:
growing a cell according to Claim 7, whereby said mammalian ATnov protein is
expressed; and
30 isolating said ATnov protein free of other proteins.
9. A purified polypeptide composition comprising at least 50 weight % of the
protein present as a ATnov protein or a fragment thereof.

10. A monoclonal antibody binding specifically to an ATnov protein.
11. A non-human transgenic animal model for ATnov gene function wherein said
5 transgenic animal comprises an introduced alteration in an ATnov gene.
12. The animal model of claim 11, wherein said animal is heterozygous for said introduced alteration.
- 10 13. The animal model of claim 12, wherein said animal is homozygous for said introduced alteration.
14. The animal model of claim 12, wherein said introduced alteration is a knockout of endogenous ATnov gene expression.
- 15 15. An isolated nucleic acid probe comprising an ATnov 3 sequence polymorphism, as part of other than a naturally occurring chromosome.
16. A nucleic acid probe according to Claim 15, wherein said probe is conjugated
20 to a detectable marker.
17. An array of oligonucleotides comprising:
two or more probes for detection of ATnov3 locus polymorphisms.

SEQUENCE LISTING

<110> Miller, Andrew
Buckler, Alan
Laubert, Boris

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Lys	Ser	Ser	Ile	Ile	His	Ile	Glu	Arg	Arg	Phe	Glu	Ile	Ser	Ser	Ser
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Ile	Val	Phe	Val	Ser	Tyr	Phe	Gly	Ser	Lys	Leu	His	Arg	Pro	Lys	Leu
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Ile	Gly	Ile	Gly	Cys	Phe	Ile	Met	Gly	Ile	Gly	Gly	Val	Leu	Thr	Ala
			100					105					110		
Leu	Pro	His	Phe	Phe	Met	Gly	Tyr	Tyr	Arg	Tyr	Ser	Lys	Glu	Thr	Asn
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Ile	Xaa	Ser	Ser	Glu	Asn	Ser	Thr	Ser	Thr	Leu	Ser	Thr	Cys	Leu	Ile
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Gly	Cys	Leu	Lys	Glu	Ser	Gly	Ser	Tyr	Met	Trp	Ile	Tyr	Val	Phe	Met
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Gly	Asn	Met	Leu	Arg	Gly	Ile	Gly	Glu	Thr	Pro	Ile	Val	Pro	Xaa	Gly
			180					185					190		
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Thr	Leu	Gly	Ser	Leu	Phe	Ser	Lys	Met	Tyr	Val	Asp	Ile	Gly	Tyr	Val
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Asp	Leu	Ser	Thr	Ile	Arg	Ile	Thr	Pro	Thr	Asp	Ser	Arg	Trp	Val	Gly
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Ala	Trp	Trp	Leu	Asn	Phe	Leu	Val	Ser	Gly	Leu	Phe	Ser	Ile	Ile	Ser
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Ser	Ile	Pro	Phe	Phe	Phe	Leu	Pro	Gln	Thr	Pro	Asn	Lys	Pro	Gln	Lys
	275					280					285				
Glu	Arg	Lys	Ala	Ser	Leu	Ser	Leu	His	Val	Leu	Glu	Thr	Asn	Asp	Glu
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 Asn Val Thr Gly Phe Phe Gln Ser Phe Lys Ser Ile Leu Thr Asn Pro
 325 330 335
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 340 345 350
 Ile Gly Ala Phe Thr Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly
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 Gln Pro Ser Ser Lys Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro
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 Lys Leu Asn Thr Val Gly Ile Ala Lys Phe Ser Cys Phe Thr Ala Val
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 Met Ser Leu Ser Phe Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn
 420 425 430
 Lys Ser Val Ala Gly Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val
 435 440 445
 Thr Ser His Arg Asp Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn
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 Cys Asp Glu Ser Gln Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr
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 Tyr Ile Ser Pro Cys Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys
 485 490 495
 Lys Pro Ile Val Phe Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu
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<212> DNA

<213> H. sapiens

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<222> (100)...(2175)

<223> Coding sequence of ATnov3.1

<221> variation

<222> (1705)...(1710)

<223> Polymorphism of 5 or 6 T residues.

<221> variation

<222> (487)...(487)

<223> Polymorphism of A or G

<221> variation

<222> (670)...(670)

<223> Polymorphism of C or T

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 Met Asp Gln Asn Gln
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cat ttg aat aaa aca gca gag gca caa cct tca gag aat aag aaa aca 162
 His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser Glu Asn Lys Lys Thr
 10 15 20

aga tac tgc aat gga ttg aag atg ttc ttg gca gct ctg tca ctc agc	210
Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala Ala Leu Ser Leu Ser	
25 30 35	
ttt att gct aag aca cta ggt gca att att atg aaa agt tcc atc att	258
Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met Lys Ser Ser Ile Ile	
40 45 50	
cat ata gaa cgg aga ttt gag ata tcc tct tct ctt gtt ggt ttt att	306
His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser Leu Val Gly Phe Ile	
55 60 65	
gac gga agc ttt gaa att gga aat ttg ctt gtg att gta ttt gtg agt	354
Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val Ile Val Phe Val Ser	
70 75 80 85	
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Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu Ile Gly Ile Gly Cys	
90 95 100	
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Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala Leu Pro His Phe Phe	
105 110 115	
atg gga tat tac agg tat tct aaa gaa act aat atc rat tca tca gaa	498
Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn Ile Xaa Ser Ser Glu	
120 125 130	
aat tca aca tcg acc tta tcc act tgt tta att aat caa att tta tca	546
Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile Asn Gln Ile Leu Ser	
135 140 145	
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Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys Gly Cys Leu Lys Glu	
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Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met Gly Asn Met Leu Arg	
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Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr Leu Gly Ile Leu Asn	
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Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe Thr Leu Gly Ser Leu	
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Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val Asp Leu Ser Thr Ile	
230 235 240 245	
agg ata act cct act gat tct cga tgg gtt gga gct tgg tgg ctt aat	882
Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly Ala Trp Trp Leu Asn	
250 255 260	

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aat ttg acc aat caa gga aaa aat att acc aaa aat gtg act ggt ttt Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys Asn Val Thr Gly Phe 310 315 320 325	1074
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tat gtc ttc aaa tac gta gag caa cag tat ggt cag cct tca tct aag Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly Gln Pro Ser Ser Lys 360 365 370	1218
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cta acc atg acc tat gat gga aat aat cca gtg aca tct cat aga gat Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val Thr Ser His Arg Asp 440 445 450	1458
gta cca ctt tct tat tgc aac tca gac tgc aat tgt gat gaa agt caa Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn Cys Asp Glu Ser Gln 455 460 465	1506
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Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu Gln Asn Arg Asn Tyr	
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Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp Ala Cys Thr Arg Lys	
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Phe Tyr Phe Phe Val Ala Ile Gln Val Leu Asn Leu Phe Phe Ser Ala	
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ctt gga ggc acc tca cat gtc atg ctg att gtt aaa att gtt caa cct	1794
Leu Gly Gly Thr Ser His Val Met Leu Ile Val Lys Ile Val Gln Pro	
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Arg Thr Tyr Asn Ser Thr Ser Phe Ser Arg Val Tyr Leu Gly Leu Ser	
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Ser Met Leu Arg Val Ser Ser Leu Val Leu Tyr Ile Ile Leu Ile Tyr	
630 635 640 645	
gcc atg aag aaa aaa tat caa gag aaa gat atc aat gca tca gaa aat	2082
Ala Met Lys Lys Lys Tyr Gln Glu Lys Asp Ile Asn Ala Ser Glu Asn	
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Gly Ser Val Met Asp Glu Ala Asn Leu Glu Ser Leu Asn Lys Asn Lys	
665 670 675	
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His Phe Val Pro Ser Ala Gly Ala Asp Ser Glu Thr His Cys *	
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<213> H. sapiens

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<222> (130)...(130)

<223> Xaa = Asp or Asn

<221> VARIANT

<222> (191)...(191)

<223> Xaa = Leu

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			20					25					30		
Ala	Leu	Ser	Leu	Ser	Phe	Ile	Ala	Lys	Thr	Leu	Gly	Ala	Ile	Ile	Met
		35					40					45			
Lys	Ser	Ser	Ile	Ile	His	Ile	Glu	Arg	Arg	Phe	Glu	Ile	Ser	Ser	Ser
	50					55					60				
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65					70					75				80	
Ile	Val	Phe	Val	Ser	Tyr	Phe	Gly	Ser	Lys	Leu	His	Arg	Pro	Lys	Leu
				85					90					95	
Ile	Gly	Ile	Gly	Cys	Phe	Ile	Met	Gly	Ile	Gly	Gly	Val	Leu	Thr	Ala
			100					105					110		
Leu	Pro	His	Phe	Phe	Met	Gly	Tyr	Tyr	Arg	Tyr	Ser	Lys	Glu	Thr	Asn
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				165					170					175	
Gly	Asn	Met	Leu	Arg	Gly	Ile	Gly	Glu	Thr	Pro	Ile	Val	Pro	Xaa	Gly
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Leu	Ser	Tyr	Ile	Asp	Asp	Phe	Ala	Lys	Glu	Gly	His	Ser	Ser	Leu	Tyr
		195				200						205			
Leu	Gly	Ile	Leu	Asn	Ala	Ile	Ala	Met	Ile	Gly	Pro	Ile	Ile	Gly	Phe
	210					215					220				
Thr	Leu	Gly	Ser	Leu	Phe	Ser	Lys	Met	Tyr	Val	Asp	Ile	Gly	Tyr	Val
225					230					235				240	
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				245					250					255	
Ala	Trp	Trp	Leu	Asn	Phe	Leu	Val	Ser	Gly	Leu	Phe	Ser	Ile	Ile	Ser
			260					265					270		
Ser	Ile	Pro	Phe	Phe	Phe	Leu	Pro	Gln	Thr	Pro	Asn	Lys	Pro	Gln	Lys
		275				280						285			
Glu	Arg	Lys	Ala	Ser	Leu	Ser	Leu	His	Val	Leu	Glu	Thr	Asn	Asp	Glu
	290					295					300				
Lys	Asp	Gln	Thr	Ala	Asn	Leu	Thr	Asn	Gln	Gly	Lys	Asn	Ile	Thr	Lys
305					310					315				320	
Asn	Val	Thr	Gly	Phe	Phe	Gln	Ser	Phe	Lys	Ser	Ile	Leu	Thr	Asn	Pro
				325					330					335	
Leu	Tyr	Val	Met	Phe	Val	Leu	Leu	Thr	Leu	Leu	Gln	Val	Ser	Ser	Tyr
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Lys Ser Val Ala Gly Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val
          435          440          445
Thr Ser His Arg Asp Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn
          450          455          460
Cys Asp Glu Ser Gln Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr
465          470          475          480
Tyr Ile Ser Pro Cys Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys
          485          490          495
Lys Pro Ile Val Phe Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu
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Gln Asn Arg Asn Tyr Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp
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Ala Cys Thr Arg Lys Phe Tyr Phe Phe Val Ala Ile Gln Val Leu Asn
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Leu Phe Phe Ser Ala Leu Gly Gly Thr Ser His Val Met Leu Ile Val
545          550          555          560
Lys Ile Val Gln Pro Glu Leu Lys Ser Leu Ala Leu Gly Phe His Ser
          565          570          575
Met Val Ile Arg Ala Leu Gly Gly Ile Leu Ala Pro Ile Tyr Phe Gly
          580          585          590
Ala Leu Ile Asp Thr Thr Cys Ile Lys Trp Ser Thr Asn Asn Cys Gly
          595          600          605
Thr Arg Gly Ser Cys Arg Thr Tyr Asn Ser Thr Ser Phe Ser Arg Val
610          615          620
Tyr Leu Gly Leu Ser Ser Met Leu Arg Val Ser Ser Leu Val Leu Tyr
625          630          635          640
Ile Ile Leu Ile Tyr Ala Met Lys Lys Lys Tyr Gln Glu Lys Asp Ile
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Asn Ala Ser Glu Asn Gly Ser Val Met Asp Glu Ala Asn Leu Glu Ser
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<211> 2360

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<222> (100)...(1729)

<223> Coding sequence ATnov3.2

<221> variation

<222> (1705)...(1710)

<223> Polymorphism of 5 or 6 T residues

<221> variation

<222> (487)...(487)

<223> Polymorphism of A or G

<221> variation

<222> (670)...(670)

<223> Polymorphism of C or T

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gct aac atc tta ttg gga gtc ata acc ata cct att ttt gca agt gga Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro Ile Phe Ala Ser Gly 375 380 385	1266
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Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys Lys Pro Ile Val Phe
                               490                               495                               500

tac aac tgc agt tgt ttg gaa gta act ggt ctc cag aac aga aat tac      1650
Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu Gln Asn Arg Asn Tyr
                               505                               510                               515

tca gcc cat ttg ggt gaa tgc cca aga gat gat gct tgt aca agg aaa      1698
Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp Ala Cys Thr Arg Lys
                               520                               525                               530

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Phe Tyr Phe Leu Leu Gln Tyr Lys Ser *
                               535                               540

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ggcatgtag gacatataat tccacatcat tttcaagggt ctacttgggc ttgtcttcaa      1989
tgtaagagt ctcatcactt gttttatata ttatattaat ttatgccaatg aagaaaaaat      2049
atcaagagaa agatatcaat gcatcagaaa atggaagtgt catggatgaa gcaaaacttag      2109
aatccttaaa taaaaataaa cattttgtcc cttctgctgg ggcagatagt gaaacacatt      2169
gttaagggga gaaaaaaagc cacttctgct tctgtgtttc caaacagcat tgcattgatt      2229
cagtaagatg ttatttttga ggagttcctg gtcctttcac taagaatttc cacatctttt      2289
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<213> H. sapiens

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<222> (130)...(130)

<223> Xaa = Asp or Asn

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<222> (191)...(191)

<223> Xaa' = Leu

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Ala Leu Ser Leu Ser Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met
                               35                               40                               45
Lys Ser Ser Ile Ile His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser
                               50                               55                               60
Leu Val Gly Phe Ile Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val
                               65                               70                               75                               80
Ile Val Phe Val Ser Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu
                               85                               90                               95
Ile Gly Ile Gly Cys Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala
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Leu Pro His Phe Phe Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn
 115 120 125
 Ile Xaa Ser Ser Glu Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile
 130 135 140

 Asn Gln Ile Leu Ser Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys
 145 150 155 160
 Gly Cys Leu Lys Glu Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met
 165 170 175
 Gly Asn Met Leu Arg Gly Ile Gly Glu Thr Pro Ile Val Pro Xaa Gly
 180 185 190
 Leu Ser Tyr Ile Asp Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr
 195 200 205
 Leu Gly Ile Leu Asn Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe
 210 215 220
 Thr Leu Gly Ser Leu Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val
 225 230 235 240
 Asp Leu Ser Thr Ile Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly
 245 250 255
 Ala Trp Trp Leu Asn Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser
 260 265 270
 Ser Ile Pro Phe Phe Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys
 275 280 285
 Glu Arg Lys Ala Ser Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu
 290 295 300
 Lys Asp Gln Thr Ala Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys
 305 310 315 320
 Asn Val Thr Gly Phe Phe Gln Ser Phe Lys Ser Ile Leu Thr Asn Pro
 325 330 335
 Leu Tyr Val Met Phe Val Leu Leu Thr Leu Leu Gln Val Ser Ser Tyr
 340 345 350
 Ile Gly Ala Phe Thr Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly
 355 360 365
 Gln Pro Ser Ser Lys Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro
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 385 390 395 400
 Lys Leu Asn Thr Val Gly Ile Ala Lys Phe Ser Cys Phe Thr Ala Val
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 Met Ser Leu Ser Phe Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn
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 Lys Ser Val Ala Gly Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val
 435 440 445
 Thr Ser His Arg Asp Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn
 450 455 460
 Cys Asp Glu Ser Gln Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr
 465 470 475 480
 Tyr Ile Ser Pro Cys Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys
 485 490 495
 Lys Pro Ile Val Phe Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu
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 <223> Polymorphism of 5 or 6 T residues

 <221> variation
 <222> (487)...(487)
 <223> Polymorphism of A or G residue

 <221> variation
 <222> (670)...(670)
 <223> Polymorphism of C or T residue.

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caacaacaaa aacatttgta tgatatctat atttcaatc atg gac caa aat caa      114
                                   Met Asp Gln Asn Gln
                                   1           5

cat ttg aat aaa aca gca gag gca caa cct tca gag aat aag aaa aca      162
His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser Glu Asn Lys Lys Thr
              10              15              20

aga tac tgc aat gga ttg aag atg ttc ttg gca gct ctg tca ctc agc      210
Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala Ala Leu Ser Leu Ser
              25              30              35

ttt att gct aag aca cta ggt gca att att atg aaa agt tcc atc att      258
Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met Lys Ser Ser Ile Ile
              40              45              50

cat ata gaa cgg aga ttt gag ata tcc tct tct ctt gtt ggt ttt att      306
His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser Leu Val Gly Phe Ile
              55              60              65

gac gga agc ttt gaa att gga aat ttg ctt gtg att gta ttt gtg agt      354
Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val Ile Val Phe Val Ser
              70              75              80              85

tac ttt gga tcc aaa cta cat aga cca aag tta att gga atc ggt tgt      402
Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu Ile Gly Ile Gly Cys
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ttc att atg gga att gga ggt gtt ttg act gct ttg cca cat ttc ttc      450
Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala Leu Pro His Phe Phe
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atg gga tat tac agg tat tct aaa gaa act aat atc rat tca tca gaa      498
Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn Ile Xaa Ser Ser Glu
              120              125              130

aat tca aca tcg acc tta tcc act tgt tta att aat caa att tta tca      546
Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile Asn Gln Ile Leu Ser
              135              140              145

ctc aat aga gca tca cct gag ata gtg gga aaa ggt tgt tta aag gaa      594
Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys Gly Cys Leu Lys Glu
              150              155              160              165
  
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gat ttc gct aaa gaa gga cat tct tct ttg tat tta ggt ata ttg aat Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr Leu Gly Ile Leu Asn 200 205 210	738
gca ata gca atg att ggt cca atc att ggc ttt acc ctg gga tct ctg Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe Thr Leu Gly Ser Leu 215 220 225	786
ttt tct aaa atg tac gtg gat att gga tat gta gat cta agc act atc Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val Asp Leu Ser Thr Ile 230 235 240 245	834
agg ata act cct act gat tct cga tgg gtt gga gct tgg tgg ctt aat Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly Ala Trp Trp Leu Asn 250 255 260	882
ttc ctt gtg tct gga cta ttc tcc att att tct tcc ata cca ttc ttt Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser Ser Ile Pro Phe Phe 265 270 275	930
ttc ttg ccc caa act cca aat aaa cca caa aaa gaa aga aaa gct tca Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys Glu Arg Lys Ala Ser 280 285 290	978
ctg tct ttg cat gtg ctg gaa aca aat gat gaa aag gat caa aca gct Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu Lys Asp Gln Thr Ala 295 300 305	1026
aat ttg acc aat caa gga aaa aat att acc aaa aat gtg act ggt ttt Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys Asn Val Thr Gly Phe 310 315 320 325	1074
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gcc atg aag aaa aaa tat caa gag aaa gat atc aat gca tca gaa aat 2082
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 665 670 675

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 His Phe Val Pro Ser Ala Gly Ala Asp Ser Glu Thr His Cys *
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 35 40 45
 Lys Ser Ser Ile Ile His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser
 50 55 60
 Leu Val Gly Phe Ile Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val
 65 70 75 80
 Ile Val Phe Val Ser Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu
 85 90 95
 Ile Gly Ile Gly Cys Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala
 100 105 110
 Leu Pro His Phe Phe Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn
 115 120 125
 Ile Xaa Ser Ser Glu Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile
 130 135 140
 Asn Gln Ile Leu Ser Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys
 145 150 155 160
 Gly Cys Leu Lys Glu Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met
 165 170 175
 Gly Asn Met Leu Arg Gly Ile Gly Glu Thr Pro Ile Val Pro Xaa Gly
 180 185 190
 Leu Ser Tyr Ile Asp Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr
 195 200 205
 Leu Gly Ile Leu Asn Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe
 210 215 220
 Thr Leu Gly Ser Leu Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val
 225 230 235 240

Asp Leu Ser Thr Ile Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly
 245 250 255
 Ala Trp Trp Leu Asn Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser
 260 265 270
 Ser Ile Pro Phe Phe Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys
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 Glu Arg Lys Ala Ser Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu
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 595 600 605
 Thr Arg Gly Ser Cys Arg Thr Tyr Asn Ser Thr Ser Phe Ser Arg Val
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 645 650 655
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 Thr His Cys
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